

Effects of Zinc and Gamma Radiation on Some Chemical Compositions of Dill Herb

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Abstract

This investigation was initiated to study the effect of foliar spray with zinc, different gamma radiation doses and their interaction treatments on chemical composition of dill. The results can be summarized as follows; pre-sowing seeds were irradiated with gamma radiation doses (0, 2, 4, 8, 16, 32 and 64 k rd) with or without zinc (150 ppm). The treatments of zinc and in most doses of gamma radiation increased the contents of chlorophyll a, b and total chlorophyll (a+b), the interaction treatments between zinc and 2, 4 and 8 k-rad of gamma radiation was superior in increasing these values. The treatments of zinc or gamma radiation up to 32 k-rad increased total carbohydrate content, the treatment of zinc combined 2 k-rad gave the highest value in this concern.

Keywords

Anethum graveolens L., Zinc, Carbohydrate, Gamma Radiation, Chlorophyll

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1. Introduction

Zinc is an essential micronutrient that acts either as a metal component of various enzymes or as a functional, structural or regulatory cofactor, and is thus associated with saccharide metabolism, photosynthesis, and protein synthesis, the synthesis of auxin, cell division, the maintenance of membrane structure and function and sexual fertilization (Marschner, 1995). Zinc deficiency has been reported to be associated with high pH soils, low extractable zinc content, calcareous soils, soils with very low organic matter and those with high available phosphorus concentrations. Zn deficiency reduces plant growth, flowering and seed production in a wide variety of plants, and in practice zinc deficiency is easy to correct by spraying or by soil application with zinc fertilizers (Takkar and Walker, 1993).Radiation processing is well established as a physical, non-thermal method to preserve various food products that involves the exposure of food products (raw or processed) to ionizing or non-ionizing radiation (Antonio et al., 2012). Irradiation of food products causes a minimal modification in the flavor, color, nutrients, taste, and other quality attributes of food (Diehl, 2002). It has been reported that under certain favorable conditions, the concentration of plant phytochemicals might be increased. These conditions contain exposure to irradiation sources, wounding, storage at low temperatures, and/or exposure to extreme temperatures (Zobel, 1996). Therefore, this study aims to study the effect of zinc and gamma radiation treatment on content of dill plant parts at vegetative and flowering developing stages.

2. Materials and Methods

2.1. Plant Material and Experimental Procedure

The experiments of this study were carried out at the Farm

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Station of National Research Centre, at Shalakan, Kalubia Governorate during the two successive seasons of 2000/2001 and 2001/2002. Seeds of dill (Anethum graveolens L.) were obtained from Medicinal and Aromatic Research Dep., Ministry of Agriculture, Egypt. Dry seeds of dill were irradiated with gamma rays doses at 2, 4, 8, 16, 32 and 64 krad before sowing. The source of irradiation is installed at the Middle Eastern Radioisotope Centre for the Arab Countries, Dokki, Giza, Egypt. Zinc EDTA (150 ppm) was applied as foliar spray at interval times of 30 and 60 days after sowing. The dill seeds were sown in 15th October in the two seasons. The experimental design of the three experiments was factorial and planned in a complete randomized block design. The concentrations of chlorophyll a, b were determined by using spectrophotometer and calculated by using Wettstein formula (Wettstein, 1957). The percentage of total carbohydrates was determined using the method described by Dubois et al. (1956).

2.2. Statical Analysis

The data of the three experiments were statistically analyzed and the differences between the means of the treatments were considered significant when they were more than least significant differences (L.S.D) at 5% level according to Steel and Torrie (1980).

3. Results and Discussion

3.1. Chlorophyll (A) Content

Data in Table(1) clearly show that leaf content of chlorophyll (a) was increased by gamma radiation, spraying zinc and their combination at vegetative stage in the two seasons in comparison to control ones. With regard to vegetative stage, it is clear that combined treatment of 64 k-rad + zinc gave the highest value (0.0838mg/100mg leaf fresh weight) in the first season, with 64 k-rad alone giving the highest value (0.0842 mg/100mg leaf fresh weight) in the second season. Spraying plants with Zn alone produced the highest values of chlorophyll (a) (0.5898 and 0.6082 mg/100mg leaf fresh weight) at flowering stage in the two seasons, respectively. Also, leaf content of chlorophyll (a) was decreased by using gamma radiation at flowering stage in both seasons. There was a significant effect on leaf content of chlorophyll (a) between the interaction treatments at flowering stage in the second season. Application of zinc treatment to dill plants significantly increased leaf content of chlorophyll (a) at both stages in the two seasons, exception those of vegetative stage in the second season; whereas, this increment was not significant. It is known that the changes in chlorophyll content are associated with the changes in chloroplasts (Preil, 1985). The important factors which control chloroplast

differentiation are: (1) genetic information present in plastids which contain the chloroplast DNA (Bidwell, 1979), (2) cytokinins which have been shown to control chloroplast differentiation independently of their action on cell division (Laloue, 1978) and (3) inorganic salts (magnesium, iron, copper, potassium and ammonium salts) which play important roles in the chlorophyll synthesis or metabolism (Bidwell, 1979). Accordingly, the effect of gamma rays doses which resulted in high amount of the leaf chlorophyll pigments in the present work can be attributed to enhancement of chloroplast differentiation and stimulation of the plastid division. Also, gamma-rays may increase endogenous cytokinins and nutrient components utilization which reflected on an increase in the leaf chlorophyll content. In this respect, Preill (1985) stated that changes in leaf chlorophyll were caused by genetic changes in chloroplast DNA and gene and/or plastid changes. Similar results were recorded by (El-Shafie et al., 1987; Eid et al., 1991; Zaharia et al., 1991; Misra, 1992; El-Esawy, 1995; Hussein et al., 1995; 1998; Ahmed, 2002; Attia, 2002; Kassem, 2002; Khalil et al., 2002; Hassanein et al., 2003).

3.2. Chlorophyll (B) Content

From the Table (2) it can be noticed that the treatments caused a positive effect on chlorophyll (b) content at both stages in the two seasons compared to control in most cases. Zinc fertilizer significantly increased chlorophyll (b) at both stages in the two seasons. In this concern, the highest content of chlorophyll (b) in the leaves was obtained by irradiated at 2 k-rad (0.0788 mg/100 mg leaf fresh weight) at vegetative stage in the first season, and (0.0767 mg/100 mg leaf fresh weight) in the second season when plants treated at 2 k-rad + zinc. In addition, spraying plants with zinc gave the highest values of chlorophyll (b) content (0.5260 and 0.5333 mg/100mg leaf fresh weight) in the flowering stage and in both seasons, respectively. Similar results were recorded by (El-Shafie et al., 1987; Eid et al., 1991; Zaharia et al., 1991; Misra, 1992; El-Esawy, 1995; Hussein et al., 1995; Mohamed, 1998; Ahmed, 2002; Attia, 2002; Kassem, 2002; Khalil et al., 2002; Hassanein et al., 2003).

3.3. Total Chlorophyll (a+b) Content

From the given data in Table (3) it can be seen that leaf content of chlorophyll (a+b) as influenced by gamma radiation, spraying plants with zinc and their interaction, behaved partly the same tendency of chlorophyll (a) or (b) as shown in Tables (1 and 2). In briefly, it can be noticed that the highest content of total chlorophyll (a+b) in vegetative stage was shown by 2 k-rad + zinc (0.1571 and 0.1565 mg/100mg leaf fresh weight) in the first and second seasons, respectively. Application of zinc significantly increased total

chlorophyll (a+b) content at both stages in the two seasons. In addition, zinc application alone, gave the highest values of chlorophyll (a+b) (1.1159 and 1.1416 mg/100 mg leaf fresh weight) in the flowering stage and in the two seasons, respectively. The above mentioned results agreed with those

obtained by (El-Shafie et al., 1987; Misra, 1992; Hussein et al., 1995; Yan and Wen, 1996; Mostafa et al., 1997; Nakhlla, 1998; Ahmed, 2002; Attia, 2002; Khalil et al., 2002;Badr et al., 2005).

 Table 1. Effect of foliar spray with zinc, gamma radiation and their interaction treatments on the chlorophyll (a) content (mg/100mg fresh leaves) of dill plants during the two seasons (2000/2001 and 2001/2002).

Zinc (ppm)	First Season							
Gamma radiation	Vegetative stage (90	days after sowing)		Flowering stage (180 days after sowing)				
(k-rad)	Zn (0 ppm)	n (0 ppm) Zn (150 ppm) Mean		Zn (0 ppm)	Zn (150 ppm)	Mean		
0	0.0696	0.0727	0.0711	0.4920	0.5898	0.5409		
2	0.0722	0.0808	0.0765	0.5050	0.5718	0.5384		
4	0.0701	0.0739	0.0720	0.4715	0.5103	0.4909		
8	0.0707	0.0783	0.0745	0.5076	0.5213	0.5144		
16	0.0771	0.0778	0.0774	0.4739	0.4990	0.4864		
32	0.0766	0.0795	0.0780	0.4969	0.4990	0.4979		
64	0.0836	0.0838	0.0837	0.5073	0.5145	0.5109		
Mean	0.0641	0.0781		0.4934	0.5293			
	Zinc = 0.0038 Zinc = 0.0211							
L.S.D. at 5%	Radiation $= 0.0071$			Radiation $=$ N.S				
	Interaction $=$ N.S			Interaction =N.S				
			Second	Season				
0	0.0701	0.0711	0.0706	0.4770	0.6082	0.5426		
2	0.0731	0.0798	0.0764	0.5106	0.5655	0.5380		
4	0.0752	0.0770	0.0761	0.4852	0.5180	0.5016		
8	0.0720	0.0755	0.0737	0.4980	0.5109	0.5044		
16	0.0795	0.0811	0.0803	0.4849	0.5002	0.4925		
32	0.0784	0.0797	0.0790	0.4899	0.5016	0.4957		
64	0.0842	0.0840	0.0841	0.5132	0.5073	0.5102		
Mean	0.0760	0.0783		0.4941	0.5302			
	Zinc = N.S			Zinc = 0.0183				
L.S.D. at 5%	Radiation = N.S			Radiation $= 0.0342$	adiation $= 0.0342$			
	Interaction = N.S			Interaction =0.0484				

 Table 2. Effect of foliar spray with zinc, gamma radiation and their interaction treatments on the chlorophyll (b) content (mg/100 mg fresh leaves) of dill plants during the two seasons (2000/2001 and 2001/2002).

Zinc(ppm)	First Season							
Gamma radiation	Vegetative stage (90 days after sowing)			Flowering stage (180 days after sowing)				
(k-rad)	Zn (0 ppm)	Zn (150 ppm)	Mean	Zn (0 ppm)	Zn (150 ppm)	Mean		
0	0.0528	0.0698	0.0613	0.4001	0.5260	0.4630		
2	0.0788	0.0763	0.0775	0.4515	0.5146	0.4830		
4	0.0705	0.0705	0.0705	0.3911	0.4749	0.4330		
8	0.0459	0.0751	0.0605	0.4306	0.5060	0.4683		
16	0.0721	0.0556	0.0638	0.3901	0.4219	0.4060		
32	0.0558	0.0741	0.0649	0.3531	0.4631	0.4081		
64	0.0682	0.0709	0.0695	0.4497	0.4252	0.4374		
Mean	0.0634	0.0703		0.4094	0.4759			
	Zinc = 00048			Zinc = 0.0306				
L.S.D. at 5%	Radiation = N.S			Radiation = N.S				
	Interaction = 0.0127 Interaction = N.S							
			Second	l Season				
0	0.0479	0.0697	0.0588	0.3614	0.5333	0.4473		
2	0.0657	0.0767	0.0712	0.4657	0.5224	0.4940		
4	0.0673	0.0705	0.0689	0.3161	0.4739	0.3950		
8	0.0427	0.0741	0.0584	0.4638	0.4895	0.4766		
16	0.0570	0.0603	0.0586	0.4051	0.4394	0.4222		
32	0.0555	0.0751	0.0653	0.3806	0.5047	0.4426		
64	0.0657	0.0713	0.0685	0.4607	0.3927	0.4267		
Mean	0.0574	0.0711		0.4076	0.4794			
	Zinc = 0.0071			Zinc = 0.0308				
L.S.D. at 5%	Radiation = N.S			Radiation $= 0.0576$				
	Interaction = N.S			Interaction $= 0.0815$				

Zinc(ppm)	- First Season								
Gamma radiation	Vegetative stage (90	days after sowing)		Flowering stage (180 days after sowing)					
(k-rad)	Zn (0 ppm)	Zn (150 ppm)	Mean	Zn (0 ppm)	Zn (150 ppm)	Mean			
0	0.1225	0.1425	0.1325	0.8921	1.1159	1.0040			
2	0.1511	0.1571	0.1541	0.9565	1.0864	1.0214			
4	0.1406	0.1444	0.1425	0.8626	0.9852	0.9239			
8	0.1166	0.1534	0.1350	0.9382	1.0273	0.9827			
16	0.1493	0.1335	0.1414	0.8641	0.9209	0.8925			
32	0.1324	0.1536	0.1430	0.8500	0.9621	0.9060			
64	0.1518	0.1547	0.1532	0.9570	0.9397	0.9483			
Mean	0.1377	0.1484		0.9029	1.0053				
	Zinc = 0.0072			Zinc = 0.0466					
L.S.D. at 5%	Radiation $= 0.0135$			Radiation $= 0.0871$					
	Interaction $= 0.0192$			Interaction = $N.S$					
			Second	Season					
0	0.1181	0.1408	0.1294	0.8385	1.1416	0.9900			
2	0.1389	0.1565	0.1477	0.9763	1.0879	1.0321			
4	0.1425	0.1475	0.1450	0.8013	0.9919	0.8966			
8	0.1148	0.1496	0.1322	0.9619	1.0004	0.9811			
16	0.1366	0.1414	0.1390	0.8900	0.9396	0.9148			
32	0.1339	0.1549	0.1444	0.8705	1.0064	0.9384			
64	0.1499	0.1553	0.1526	0.9739	0.9000	0.9369			
Mean	0.1335	0.1494		0.9017	1.0096				
	Zinc = 0.0105			Zinc = 0.0408					
L.S.D. at 5%	Radiation = N.S			Radiation = 0.0764					
	Interaction = N.S			Interaction $= 0.1080$					

 Table 3. Effect of foliar spray with zinc, gamma radiation and their interaction treatments on the total chlorophyll (a+b) content (mg/100 mg fresh leaves) of dill plants during the two seasons (2000/2001 and 2001/2002).

3.4. Total Carbohydrate Percentage

3.4.1. In the Aerial Parts

Data in Table (4) clearly revealed that, spraying plants with zinc alone induced significant increase of total carbohydrate percent in dill over control in both stages and seasons. Gamma radiation led to a considerable simulative effect on total carbohydrate percent in the flowering stage of both seasons comparison to that of control plants. On the other hand, gamma rays (32 and 64 k-rad) tended to a decrease in total carbohydrate percent of dill herb in the vegetative stage during the two seasons in comparison to control. Differences between interaction treatments were not significant at flowering stage in both seasons, while, there were significant differences between the interaction treatments of zinc and gamma radiation at vegetative stage in the two seasons. The largest percent of total carbohydrate was found by combined treatment of 8 k-rad + zinc (30.12 and 29.84%) at vegetative stage in both seasons, respectively. In the flowering stage, plants irradiated at 2 k-rad + zinc gave the highest values as follows: herb (23.18, 22.70%), umbels (23.52, 23.01%) and leaves + stems (22.84, 22.40%) in the first and second seasons, respectively. the effects of different radiation doses on enzyme activity of α-amylase. Increased amaylase activity which precedes differentiation may be necessary for increased mobilization of carbohydrate reserves concomitant with high synthetic activities which occur during organogensis (Thrope and Meier, 1972). The stimulatory effect of zinc on carbohydrate content would be attributed to its role in activation of the enzymes responsible for photosynthesis, biosynthesis and transformation of carbohydrates, regulation of sugars and starch formation, (Marschner, 1995). Similar results were recorded by (El-Sherbeny and Abou-Zied, 1986; Hussein et al., 1990; El-Shafie et al., 1990; Abou-Leila et al., 1994; Cai and Han, 1994; Abou-Zied et al., 1996; Mohamed, 1998; Khalil et al., 2001; Attia, 2002; Khalil et al., 2002).

3.4.2. In the Roots

Results in Table (5) indicate that all treatments were affected on total carbohydrate percent in dill roots during both stages in both seasons. Increasing gamma doses up to 16 and 32 k-rad increased total carbohydrates percent at both stages in the first and second seasons, respectively, with significant increases during both stages and the two seasons with gamma ray at 2 k-rad in comparison to unirradiated plants. The highest values of total carbohydrate percent (25.73 and 25.34% in the vegetative stage and 20.87, 20.35% in the flowering stage at both seasons, respectively) were found by plants irradiated at 2 k-rad + zinc.Regarding the effect of zinc on total carbohydrate percent, data in Table (5) show that, application of zinc significantly increased total carbohydrate percent at vegetative stage in both seasons, whereas, these increments were significant at flowering stage in second season. with 64 k-rad alone giving lower values of total carbohydrate percent in dill roots at both stages in the two seasons. Similar results were recorded by Ahmed (2002) on garlic.

		First Season											
Zinc(ppm) Gamma)	Vegetative stage (90 days after sowing)			Flowering stage (180 days after sowing)								
radiation		Herb			Umbels			Leaves + Stems			Herb		
(K-rad)		Zn (0	Zn (150	Mean	Zn	Zn (150	Mean	Zn	Zn	Moon	Z_n (0 ppm)	Zn	Mean
		ppm)	ppm)	Wittan	(0 ppm)	ppm)	wittan	(0 ppm)	(150 ppm)	Witan		(150 ppm)	witan
0		26.81	27.01	26.91	17.15	20.01	18.58	18.73	19.67	19.20	17.94	19.84	18.89
2		27.19	27.34	27.26	21.74	23.52	22.63	20.88	22.84	21.86	21.31	23.18	22.24
4		25.78	28.57	27.17	21.72	21.93	21.82	20.14	21.55	20.84	20.93	21.74	21.33
8		26.40	30.12	28.26	20.20	21.92	21.06	19.26	21.94	20.60	19.73	21.93	20.83
16		25.20	29.39	27.29	19.16	21.64	20.40	19.07	21.18	20.12	19.12	21.41	20.26
32		24.47	27.40	25.93	19.93	23.10	21.51	19.19	21.08	20.13	19.56	22.09	20.82
64		20.69	24.51	22.60	18.40	21.55	19.97	18.54	19.96	19.25	18.47	20.75	19.61
Mean		25.22	27.76		19.75	21.95		19.40	21.17		19.58	21.56	
LCD of	. +	Zinc = 0.79			Zinc = 0.52			Zinc = 0.40			Zinc = 0.43		
L.S.D. at	ι	Radiation = 1.48			Radiation $= 0.97$			Radiation $= 0.75$			Radiation $= 0.81$		
570		Interaction $= 2.09$			Interaction $= 1.38$			Interaction = N.S			Interaction $=$ N.S		
							Secon	d Season					
0		25.80	25.91	25.85	18.15	20.06	19.10	18.93	20.70	19.81	18.54	20.38	19.46
2		26.68	27.57	27.12	22.16	23.01	22.58	21.04	22.40	21.72	21.60	22.70	22.15
4		25.94	28.16	27.05	20.68	21.45	21.06	19.95	21.80	20.87	20.31	21.63	20.97
8		25.47	29.84	27.65	20.11	22.54	21.32	19.47	21.82	20.64	19.79	22.18	20.98
16		25.62	29.80	27.71	20.55	21.32	20.93	19.77	21.18	20.47	20.16	21.25	20.70
32		21.94	28.30	25.12	21.44	22.50	21.97	19.97	21.29	20.63	20.71	21.89	21.30
64		20.56	24.50	22.53	19.60	20.43	20.01	19.27	19.84	19.55	19.44	20.13	19.78
Mean		24.57	27.72		20.38	21.61		19.77	21.29		20.07	21.45	
ISD of	+	Zinc = 0.66			Zinc = 0.52			Zinc = 0.33			Zinc = 0.37		
L.S.D. at	ι	Radiation $= 1.25$			Radiation $= 0.98$			Radiation = 0.63			Radiation $= 0.70$		
5%		Interaction $= 1.76$			Interaction $=$ N.S			Interaction $= N.S$			Interaction $=$ N.S		

 Table 4. Effect of foliar spray with zinc, gamma radiation and their interaction treatments on the total carbohydrate content (%) of dill plants during the two seasons (2000/2001 and 2001/2002).

 Table 5. Effect of foliar spray with zinc, gamma radiation and their interaction treatments on the total carbohydrate content (%) in the roots of dill plants during the two seasons (2000/2001 and 2001/2002).

Zinc(ppm)	First Season								
Gamma radiation	Vegetative stage (9	0 days after sowing)		Flowering stage (180 days after sowing)					
(k-rad)	Zn (0 ppm)	Zn (150 ppm)	Mean	Zn (0 ppm)	Zn (150 ppm)	Mean			
0	22.29	23.48	22.88	18.99	20.00	19.49			
2	23.66	25.73	24.69	19.92	20.87	20.39			
4	22.46	24.04	23.25	18.53	19.24	18.88			
8	22.60	24.65	23.62	20.22	18.78	19.50			
16	22.96	24.65	23.80	19.75	19.85	19.80			
32	22.38	22.40	22.39	17.68	18.55	18.11			
64	19.53	20.37	19.95	17.92	17.60	17.76			
Mean	22.26	23.61		19.00	19.27				
L.S.D. at 5%	Zinc = 0.66 Radiation = 1.25 Interaction = N.S			Zinc = N.S Radiation =0.80 Interaction =1.14					
			Second Season						
0	21.85	22.30	22.07	17.87	18.75	18.31			
2	23.19	25.34	24.26	19.49	20.35	19.92			
4	21.87	24.73	23.30	18.21	19.98	19.09			
8	22.00	23.22	22.61	19.33	19.19	19.26			
16	22.62	23.15	22.88	19.06	19.49	19.27			
32	22.56	23.12	22.84	17.97	19.03	18.50			
64	19.62	20.68	20.15	17.78	18.54	18.16			
Mean	21.95	23.22		18.53	19.33				
	Zinc = 0.74			Zinc = 0.49					
L.S.D. at 5%	Radiation = 1.39			Radiation = 0.92	Radiation = 0.92				
	Interaction =N.S			Interaction =N.S					

4. Conclusion

The recorded results showed that in the two seasons, the interaction treatment between zinc and gamma radiation doses (2, 4 and 8 k-rad) recorded higher values for chlorophyll (a), (b), total chlorophyll (a+b). Also, the most effective treatment 2 k-rad + zinc was the best for total carbohydrate in different plant parts.

References

- Abou-Leila, B.H.; M.S. Aly and N.F. Abdel-Hady (1994). Effect of foliar application of GA and Zn on *Ocimum basilicum* L. grown in different soil type. Egypt. J. Physiol. Sci., 18, No. 2, pp. 365-380.
- [2] Abou-Zeid, E.N.; N.Y. Nagiub; M.S. Hussein and S.E. EL-Sherbeny (1996). Response of *Ricinus communis* to gamma irradiation and sulphur fertilizer. Egypt. J. Appl. Sci., 11 (4): 117-130.
- [3] Ahmad, O.O.M. (2002). Physiological studies on garlic crop. M.Sc. Thesis, Fac. of Agric., Zagazig Univ.
- [4] Antonio, A.L., Carocho, M., Bento, A., Quintana, B., Luisa Botelho, M. and I.C. Ferreira (2012). Effects of gamma radiation on the biological, physico-chemical, nutritional and antioxidant parameters of chestnuts– A review. Food Chem. Toxicol., 50:3234-3242.
- [5] Attia, M.M.A.A. (2002). Growth and composition of marigold as affected by chemical and organic fertilization in the presence of copper and zinc. Ph.D. Thesis, Fac. of Agric., Cairo Univ.
- [6] Badr, M.; B.A. Abdel-Maksoud and S.S. Omer (2004). Growth, flowering and induced variability in *Gomphrena globosa* L. plant grown from dry and water-soaked seeds treated with gamma-rays. Alex. J. Agric. Res., 49 (1): 49-70.
- [7] Bidwell, R.G.S. (1979). Plant Physiology. Second Edition, Machmillan Publishing Co., Inc. New York.
- [8] Diehl, J.F. (2002). Food irradiation-past, present and future. Radiat. Phys. Chem., 63:211-215.
- [9] Dubois, M.; K.A. Gilles; J.K. Hamilton; P.A. Bebra and F. Smith (1956). Colorimetric methods for determination of sugars and related substances. Analyt. Chem., 28 (3): 350.
- [10] Cai, W. and F.X. Han (1994): Effect of zinc on carbon and nitrogen metabolism in tea plant (*Camellia sinensis* L.). Scientia Agric. Sinica, 27 (2): 72-77.
- [11] Eid, S.M.; N.S. Shafshak and F.A. Abou-Sedera (1991). Effect of potassium fertilization and foliar application of certain micro-nutrients combinations on growth, yield and chemical composition of garlic plants. Ann. Agric. Sci., Moshtohor, 29 (2): 981-993.
- [12] El-Esawy, M.M.A. (1995). Effect of radiation and gibberellic acid on the growth and flowering of gladiolus corms. Ph.D. Thesis, Fac. of Agric., Ain Shams Univ.
- [13] El-Shafie, S.A.; F.A. Mohamed; El-Kholy and M.M. Abdel-Baky (1987). Physiological response of carnation plant to

gamma rays. 12th Int. Conf. for Statist., Comp. Sci., Social and Demo. Res., Cairo, 1987. pp. 45-56.

- [14] El-Shafie, S.A.; M.M. Mazrou and S.A. El-Sayed (1993). The influence of pre-sowing laser-irradiation on the growth, productivity and some chemical constituents of *Ammi visnaga* L. plants. Menofiya J. Agric. Res., vol. 18 No. 4 (2): 2591-2604.
- [15] El-Sherbeny, S.E. and E.N. Abou-Zeid (1986). A preliminary study on the effect of foliar micro-elements on growth and chemical constituents in *Foeniculum capillacum*. Bull. NRC. Egypt, 11, 606-612.
- [16] Hassanein, R.A.M. (2003). Effect of some amino acids, trace elements and irradiation on fennel (*Foeniculum vulgare* L.). M.Sc. Thesis, Fac. of Agric., Cairo Univ.
- [17] Hussein, M.S.; S.E. El-Sherbeny and N.Y. Nagiub (1995). The effect of γ-radiation and manganese application on growth and chemical constituents of Daturametel L. Egypt J. Physiol. Sci., 19, No. 1-2, pp. 241-254.
- [18] Kassem, A.H.M.M. (2002). Effect of planting distances and some trace elements on rosemary plant. Ph.D. Thesis, Fac. of Agric., Cairo Univ.
- [19] Khalil, M.Y.; N.Y. Nagiub and S.E. El-Sherbeny (2002). Response of *Tagetes erecta* L. to compost and foliar application of some micro-elements. Arab Univ. J. Agric. Sci., Ain Shams Univ., Cairo, 10 (3): 939-964.
- [20] Khalil, M.Y.; S.E.El-Sherbeny and M.S. Hussein (2001). Growth, yield and chemical constituents of some medicinal plants in relation to gamma irradiation. Egypt J. Hort., 28, (3): 355-369.
- [21] Laloue, M. (1978): Functions of cytokinins. Phil. Trnas R. Soci., Land B. 284: 449-457.
- [22] Marschner, H. (1995). Mineral Nutrient of Higher Plants. Second Edition. Academic Press Limited. Harcaurt Brace & Company, Publishers. London, pp. 347-364.
- [23] Misra, A. (1992). Effect of zinc stress in Japanese mint as related to growth, photosynthesis, chlorophyll contents and secondary plant products- the monoterpenes. Photosynthetica, 26 (2): 225-234.
- [24] Mostafa, M.M.; E.H. El-Haddad and M.A. Amer (1997). Effectiveness of foliar nutrition with some micro-elements of chrysanthemum plants. Alex. J. Agric. Res., 42 (1): 81-93.
- [25] Nakhlla, F.G. (1998): Zinc spray on navel orange in newly reclaimed desert areas and its relation to foliar IAA level and fruit drop. Bull. Fac. Agric., Univ. Cairo, 49 (1): 69-88.
- [26] Preil, W. (1985). In vitro propagation and breeding of ornamental plants: advantage and disadvantage of variability. Genetic Manipulation in plant Breeding, Berlin (west). Inter. Symp. Org. by Eucarpia, pp. 55 No. 42.
- [27] Steel, R.G.D. and S.H. Torrie (1980): Principales and Procedure of Statistics. Second Edition, McGrow Hill Inc.
- [28] Takkar, P.N. and C.D. Walker (1993). The distribution and correction of zinc deficiency. In Zinc in Soil and Plants Proceddings of the International Symposium on "Zinc in Soil and Plants" held at the University of Western Australia, 27-28 Septemper, 1993. Ed. A.D. Robson, Klumer Academic Publishers, pp. 151-165.

- [29] Thorpe, T.A. and D.D. Meier (1972). Starch metabolism, respiration and shoot formation in tobacco callus cultures. Physiologia Plantarum, 27: 365-9.
- [30] Yan, H.W. and X.Y. Wen (1996). Physiological effects of Cu and Zn on the development and physiological metabolism of tea plants. V. Effect of interaction between Cu and Zn on the growth of tea plants. J. of Tea Sci., 16 (2): 99-104.
- [31] Wettstein, D.V. (1957). Chlorophyll- lethale under Submikroskopischeformivechrel der plastieden. Exptl. Celi. Res., (12): 427-433.
- [32] Zaharia, D.; D. Popa and V. Bercea (1991). Effect of gamma irradiation on the seed germination and biosynthesis of assimilating pigments in several ornamental plants. BuletinulInstitutului Agronomic cluj Napoca Seria Agricultura, 44 (1): 107-114.
- [33] Zobel, A.M., editor. (1996). Coumarins in fruit and vegetables. Proceeding-Phytochemical Society of Europe; Oxford Univesity Press INC.