

Efficacy of Certain Plant Extracts as Safe Fungicides Against Phytopathogenic and Mycotoxigenic Fungi

Abd El-Ghany T. M.^{1, 2, *}, Roushdy M. M.¹, Mohamed A. Al Abboud²

¹Botany and Microbiology Department, Faculty of Science, AL-Azhar University, Cairo, Egypt

²Biology Department, Faculty of Science, Jazan University, Jazan, 114, Kingdom Saudi Arabia

Abstract

The efficiency of different natural plant extracts and of the chemical fungicide Micronite was carried out to determine their effects on the soil fungi particularly phytopathogenic and mycotoxigenic fungi *Fusarium oxysporum*, *Alternaria alternata* and *Aspergillus flavus*. *In vitro* studies were carried out to test the antifungal activity of 4 plant extracts; performed with either cold distilled water. The results revealed that plants extracts had a strong antifungal activity with significant inhibition on the growth of the all tested fungi. Extracts of *Azadirachta indica* and *Jatropha curcas* were the most effective to inhibit the growth of the tested fungi. On the other hand, the chemical fungicide was more efficient than the natural compounds. Different concentrations of plant extract of *A. indica* and of chemical fungicide were studied on the growth of *Aspergillus flavus* and *Alternaria alternata*. Findings from this study confirmed that plant extracts can be used as natural fungicides to control pathogenic fungi, thus reducing the dependence on the synthetic fungicides. *Azadirachta indica* extract, which was found to be the most efficient extract, might be a promising agent for controlling these fungi.

Keywords

Fungi, Bioevaluating, Fungicide, Fungi, Mycotoxigenic, Phytopathogenic

Received: March 23, 2015 / Accepted: April 4, 2015 / Published online: April 6, 2015

@ 2015 The Authors. Published by American Institute of Science. This Open Access article is under the CC BY-NC license.

<http://creativecommons.org/licenses/by-nc/4.0/>

1. Introduction

To avoid the hazardous effects of chemicals, natural products of some plants have been used to control plant disease (Rahber-Bhatti, 1986; Bowers and Locke, 2000; Momin *et al.*, 2001). Development of safer anti-fungal agents such as plant extracts to control phytopathogens in agriculture was reported in recent years (Imtiaj *et al.*, 2005; Tumen *et al.*, 2013). The essential oils and their constituents have been found effective as antifungal agent (Daferera *et al.*, 2000; Sridhar *et al.*, 2003). Several reports have been made on the fungicidal properties of neem oil (Kazmi *et al.*, 1995). Locke (1995) reported that in field *Alternaria alternata*, *Aspergillus niger* and *Fusarium oxysporum* has been completely controlled by using 2-10% neem oil. It is observed that

mustard seed oil also showed antifungal activity (Dhingra *et al.*, 2004). Houghton *et al.*, (2006) reported antifungal activity of asafoetida against *Microsporeum gypseum* and *Trichophyton interdigitale*. Thyagaraia & Hosono (1996) also studied the inhibition effect of asafoetida on *Rhizopus sporus*, *Mucor dimorphosporous*, *Penicillium commune* and *Fusarium solani*. Extracts from plants such as garlic (*Allium sativum*) (Obagwu and Korsten, 2003), *Azadirachta indica*, *Moringa oleifera* (Adandonon *et al.*, 2006), *Ferula communis* and *Ditrichia viscosa*, *Juniperus communis* (Menghani and Sharma 2012) have been tested on many other soil borne fungi. Alkhail (2005) showed that extracts of *Allium sativum*, *Azadirachta indica* and *Eugenia caryophyllus* presented remarkable biological activity when tested against fungi viz., *F. oxysporum*, *Botrytis cinerea*.

* Corresponding author

E-mail address: tabdelghany@yahoo.com (T. M. Abd El-Ghany)

The antimicrobial and antitoxin properties of some plants, herbs, and their components have been documented since the late 19th century (Saadabi, 2006, Fawzi *et al.* 2009, Zaker and Mosallanejad 2010, Abdulghaffar *et al.* 2010; Abdel Ghany and Hakamy 2014). These natural plants involve garlic, lemon grass, datura, acacia, a triplex, ginger, black seed, neem, basil, eucalyptus, *Juniperus procera*, alfalfa and basil (Omar and Abd-El-Halim, 1992; Aly *et al.*, 2000; Aly and Bafiel, 2008; Abdel Ghany 2014). They are safer to human and the ecosystem than the chemical antifungal compounds, and can easily be used by the public who used them for thousands of years to enhance flavor and aroma of foods as well as its economic value (Shelef, 1983). An emerging alternative to random chemical synthesis is the study and exploitation of naturally occurring products with fungicidal properties. Plants produce an enormous array of secondary metabolites, and it is commonly reasoned that a significant part of this chemical diversity serves to protect plants against plant pathogens. A problem with plant-produced compounds as potential fungicides is that in the natural state, they are generally only weakly active compared to synthetic fungicides. The efficiency of different natural plant extracts and chemical fungicide (Micronite) was carried out to determine their effects on the soil fungi particularly phytopathogenic and mycotoxigenic fungi *Fusarium oxysporum* and *Alternaria alternata* and *Aspergillus flavus*. *In vitro* studies were carried out to test the antifungal activity of 4 plant extracts; performed with either cold distilled water. Therefore, this work aimed to study the evaluating the antifungal activity of natural compounds of plant extracts with comparing of chemical fungicide (Micronite).

2. Materials and Methods

2.1. Plant samples and Fungal Culture Used

Leaves of *Azadirachta indica*, *Jatropha curcas*, *Ricinus communis* and *Allium sativum* were dried and ground into a fine powder in an electric grinder and extracted with distilled water. The extract was added for growth medium for testing the antifungal properties against fungal species *Aspergillus flavus*, *Alternaria alternata*, *Fusarium oxysporum*, *Rhizopus stolonifer* and *Cladosporium herbaru*.

2.2. Poisoned Food Technique

Potato dextrose agar medium (PDA) with different concentration plant extracts of the test plants were prepared. About 25 ml of the growth medium was poured into each petri-dish and allowed to solidify. Five mm disc of 5-day-old culture of the test fungi was placed at the center of the petridish and incubated at 27°C for 7 days, the growth was measured in millimeter. For each treatment three replicates

were maintained. PDA medium with micronite (chemical fungicide) was used as a positive control. PDA medium without the methanolic extract served as control. The fungitoxicity on the extracts in terms of percentage inhibition of mycelia growth was calculated by using the formula: % inhibition = $(dc - dt) / dc \cdot 100$; Where, dc = average increase in mycelial growth in control, dt = Average increase at each treatment (Singh and Tripathi, 1999).

2.3. Chemical Fungicide

Micronite: Micronite containing active ingredient 80% sulfur (Fig. 1) produced by National company for agrochemical production used as fungicide 25g / 10 liter water.

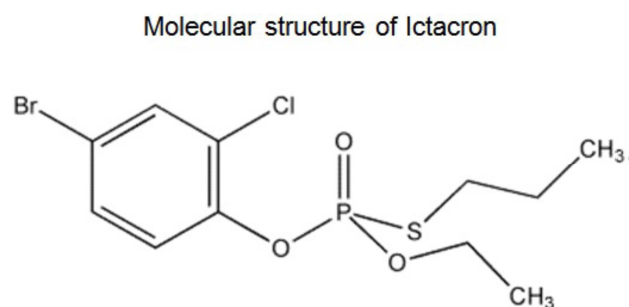


Fig. 1. Chemical structure of Micronite.

3. Result and Discussion

The results revealed that plants extracts had a strong antifungal activity with significant inhibition on the growth of the all tested fungi. Extracts of *Azadirachta indica* and *Jatropha curcas* were the most effective to inhibit the growth of the tested fungi. On the other hand, the chemical fungicide was more efficient than natural compounds. Garlic plant extract in this study showed antifungal activity (Table 1) The results of this study corresponds with work done by William (2008) who reported that sprays made from aqueous garlic extracts have antibiotic and antifungal properties and will suppress a number of plant diseases, including powdery mildew on cucumbers and to some extent black spot on roses. Similar results were reported by Slusarenko *et al.* (2008) who tested the effectiveness of garlic juice against a range of plant pathogenic bacteria, fungi and oomycetes *in vitro*. The effects of the antifungal compounds may be on spore germination leading to its inhibition or may be due to effect of these compounds on the cell wall altering its permeability (William, 2008). Our results showed the antifungal activity of *Azadirachta indica* extract. Mycelial growth of various species of *Fusarium* was inhibited by the plant extracts of *Allium cepa* (Patel, 1989), *Cassia nodosa* (Reddy and Reddy, 1987); *Azadirachta indica* (Eswaramoorthy *et al.*, 1989);, *Allium sativum* and *Sapindus trifoliata* (Gohil and Vala, 1996); Neem seed extract (Gour and Sharmaik, 1998),

Eucalyptus amygdalina, (Bansal and Gupta, 2000). In accordance with the above reports, in the present study, 87.76 % inhibition of mycelial growth of *Aspergillus flavus* of leaf extracts of *Azadirachta indica* and 79.09 % inhibition of *Alternaria alternata* of mycelial growth (Table 1). Different concentrations of plant extract of *Azadirachta indica* and chemical fungicide were studied on the growth of *Aspergillus flavus*, *Fusarium oxysporum* and *Alternaria alternata* (Fig.2). As well as increasing in the concentration of *Azadirachta indica* extract and chemical fungicide inhibition of *A. flavus* and *Alternaria alternata* increased (Table 2 and 3). The plant extract of *Azadirachta indica* exhibited effect on *Alternaria alternata* spores causing decreasing in the size and rate of sporogenesis. This result is agree with result obtained by Mondall *et al.* (2009) where the crude aqueous and alcoholic leaf extracts of *Azadirachta indica* was more effective in inhibitions of growth of the fungi *Aspergillus* in comparison to inhibitory effects on *Rhizopus* growth in the artificial culture medium. Alam *et al.* (2004) tested five plant extracts against conidial germination of *Fusarium oxysporum* and reported that the extract of *Calotropis procera* showed high inhibitory effect. Effect of plant extracts on conidial germination, mycelial growth and sporulation of *Aspergillus flavus*, *A. niger* and *A. fumigatus*

were examined (Locke, 1995; Bansal and Gupta, 2000; William, 2008) where, *Lowsonia inermis* inhibited conidial germination of *A. flavus* and *A. fumigatus*, while *A. niger* was mostly inhibited by *A. indica*. On the other hand chemical fungicide exhibited more effect on spores causing very reductin in the size and number of cells per conidiospores. Findings from this study confirmed that plant extracts can be used as natural fungicides to control pathogenic fungi, thus reducing the dependence on the synthetic fungicides. *Azadirachta indica* extract, which was found to be the most efficient extract, might be a promising material for controlling these fungi. Mixture of chemical fungicide with *A. indica* extract (Table 4) increased the antifungal activity against tested fungi.

4. Conclusion

The results of the present study revealed that *J. curcas*, *A.indica*, *A. sativum* and *R. communis* extracts have been emerged as safe alternatives to replace chemical fungicides and can be used as eco-friendly fungicides. Further work is required to increase the efficacy of these plant extracts in the field and also to determine the biologically active ingredient present in extracts as well as its mode of action.

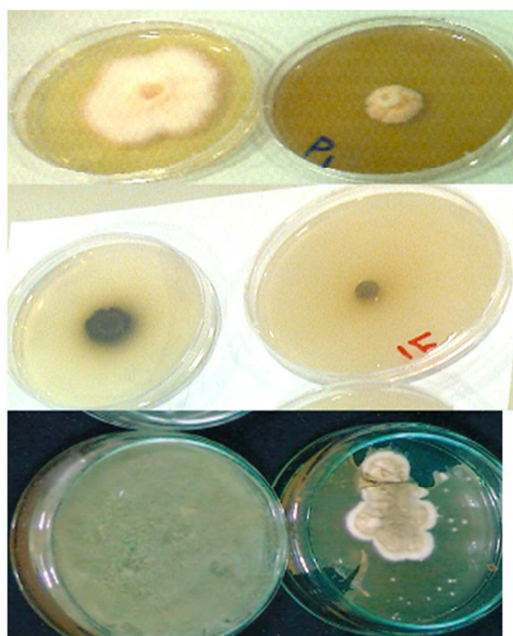


Fig. 2. Effect of *Azadirachta indica* extract on *Fusarium oxysporum*, *Alternaria alternata* and *Aspergillus flavus* From higher to lower respectefly. Left is Control without treatment; Right is treated with *Azadirachta indica* extract.

Table (1). Inhibitory effect of natural plant extracts and chemical fungicide. Data are expressed as % of fungal growth inhibition.

Chemical fungicide	Control	<i>Jatropa curcas</i>	<i>Azadirachta indica</i>	<i>Allium sativum</i>	<i>Ricinus communis</i>	Test fungi
94.98	0.00	77.08	87.76	66.8	45.9	<i>A. flavus</i>
80.76	0.00	56.80	78.76	54.87	78.77	<i>P. cyclopium</i>
88.67	0.00	67.54	49.65	43.98	48.98	<i>F. oxysporum</i>
95.00	0.00	78.00	79.09	45.89	69.32	<i>A. alternata</i>
90.45	0.00	70.60	77.08	48.78	70.45	<i>R. nigricans</i>
94.50	0.00	89.50	84.56	76.76	48.34	<i>C. herbarum</i>

Table (2). Effect of different concentrations of chemical fungicide.

Colony radius (cm)		Concentration mg %
<i>Alternaria alternata</i>	<i>Aspergillus flavus</i>	
6.3	6.5	Control
5.5	4.0	1
2.0	2.5	2
1.3	1.6	4
0.0	0.0	6

Table (3). Effect of different concentrations of *Azadirachta indica* extract.

Colony radius(cm)		Concentration mg %
<i>Alternaria alternata</i>	<i>Aspergillus flavus</i>	
6.3	6.5	Control
6.3	6.0	1
4.0	5.5	2
3.4	3.7	4
2.5	3.3	6

Table (4). Effect of different concentrations of mixture in equal amount of *Azadirachta indica* extract and Chemical fungicide.

Colony radius (cm)		Concentration mg %
<i>Alternaria alternata</i>	<i>Aspergillus flavus</i>	
6.3	6.5	Control
5.1	4.0	1
1.5	1.5	2
1.2	1.2	4
0.0	0.0	6

Acknowledgement

Thanks to Regional Center of Mycology and Biotechnology, Al Azhar University, Cairo, Egypt for providing laboratory facilities.

References

- [1] Abdel Ghany T.M. (2014). Eco-friendly role of *Juniperus procera* as safe alternative for controlling fungal growth and their secondary metabolites. The African Journal of Mycology and Biotechnology, 19 (1), 21-36.
- [2] Abdel Ghany T.M. and Hakamy O.M. (2014). *Juniperus procera* as food safe additive, their antioxidant, anticancer and antimicrobial activity against some food-borne bacteria. J. Biol. Chem. Research., 31(2): 668-677.
- [3] Abdulghaffar O.; Abbas H.; Karim M. (2010). Samira Ebrahimzadeh Antifungal Effects of Thyme, Agastache and Satureja Essential Oils on *Aspergillus fumigatus*, *Aspergillus flavus* and *Fusarium solani* Veterinary Research Forum Vol: 1, No: 2, pp. 99 – 105.
- [4] Adandonon, A., Aveling, T.A.S., Labuschagne, N., Tamo M. (2006). Biocontrol agents in combination with *Moringa oleifera* extract for integrated control of *Sclerotium*-caused cowpea damping-off and stem rot. European Journal of Plant Pathology, 10, 1-6.
- [5] Alkhail, A.A. (2005). Antifungal activity of some extracts against some plant pathogenic fungi. Pakistan Journal of Biological Sciences, 8(3), 413-417.
- [6] Alam S, Islam M R, Sarkar M A, Chowdhury A N, Alam M S and Lee M W (2004) In vitro effect of fungicides, plant extract and smoke on conidial germination of *Fusarium oxysporum* root rot pathogen of Piper beetle. Mycobiology 32 (1): 42-46.
- [7] Aly AA, Omar SA, Zayed SME, Mansour MTM, (2000). Use of saponin- containing *Atriplex nummularia* to suppress damping of cotton seedling. J. Agric. Sci. Mansura Uni., 25: 7621-7631.
- [8] Aly MM, Bafiel S, (2008). Screening for antimicrobial activity of some medicinal plants in Saudi Arabia. World conference on medical and aromatic
- [9] Bansal K R and Rajesh K G. (2000). Evaluation of plant extracts against *Fusarium oxysporum*, wilt pathogen of fenugreek, Indian. J. Phytopath., 53 (1) 107-108.
- [10] Bowers, J.H and J.C. Locke. (2000) Effect of botanical extracts on population density of *Fusarium oxysporum* in soil and control of *Fusarium* wilt in the green house. Plant Dis., 88: 300-305.
- [11] Daferera, D.J, B.N. Zirgas and M.G. Polission. (2000). GC-MS Analysis of essential oil from some Greek aromatic plants and their fungitoxicity on *Penicillium digitatum*. J. Agric. Food. Chem., 48: 2576-2581.
- [12] Dhingra, O.D., M. L.N. Costa, G.J. Silva and E.S.G. Mizubuti. (2004). Essential Oil of Mustard to control *Rhizoctonia solani* causing seedling damping off and seedling blight in nursery. Fitopatologia brasileira, 24: 683-686.
- [13] Eswaramoorthy S, Muthusamy S and Mariappan V. (1989). Neem, News letter, 6 (1) (1989) 4-5.

- [14] Fawzi, E. M., Khalil, A. A. and Afifi, A. F. (2009). Antifungal effect of some plant extracts on *Alternaria alternata* and *Fusarium oxysporum* African Journal of Biotechnology Vol. 8 (11), pp. 2590-2597 .
- [15] Gohil V P and Vala G D. (1996) .Effect of extracts of some medicinal plants on the growth of *Fusarium moniliforme*, J. Mycol. Pl. Pathol., 26 (1) 110- 111.
- [16] Gour HN, Sharmaik C (1998). Inhibition of growth, Sporulation and Phytotoxicity of *Fusarium oxysporum* fungal species Cumini, a wilt pathogen of cumin by plant extracts. J. Mycol. Plant Pathol., 2: 76- 77.
- [17] Houghton, P.J., K.M. Ismail, L. Maxia and G. Appendino. (2006). Antidermatophytic prenglated coumarins from asafoetida. *Planta Med.*, 72 DOI; 10.1055/S – 949741.
- [18] Imtiaj, A., Syed, A. R., Shahidul, A., Rehana, P., Khandaker, M. F., Sang-Beom, K., Tae-Soo, L. (2005). Effect of Fungicides and Plant Extracts on the Conidial Germination of *Colletotrichum gloeosporioides* Causing Mango Anthracnose, *Mycobiology* 33(4), 200-205.
- [19] Kazmi, S.A.R., S. Shahzad and I. Niaz. (1995). Effect of neem oil on in vitro growth of root infecting fungi. *Pak. J. Bot.*, 27(1): 217-220.
- [20] Locke, J.E. (1995). Fungi. In: *The Neem Tree, source of Unique National Products for Integrated pest Management, Medicine, Industry and Other proposes.* (Ed.): H. Schmutterer. V.C.H,Weinheim, Germany. p 118-125.
- [21] Menghani, K. and Sharma, S. K. (2012). Antimicrobial activity of *Juniperus communis* and *Solanum xanthocarpum*. *International Journal Pharmaceutical Sciences and Research*, 3(8), 2815-2818.
- [22] Momin R.A. and Nair M.G. (2001). Mosquitocidal Nematicidal and antifungal compounds from *Apium graveolens* L., seeds. *J. Agric. Food Chem.*, 49: 142-145.
- [23] Mondall , N. K.; Mojumdar., A.; Chatterje, S. K.; Banerjee, A.;Datta, J.K.; Gupta, S. (2009) Antifungal activities and chemical characterization of Neem leaf extracts on the growth of some selected fungal species in vitro culture medium. *J. Appl. Sci. Environ. Manage* Vol. 13(1) 49 – 53.
- [24] Obagwu, J., Korsten, L. (2003). Control of citrus green and blue molds with garlic extracts. *European Journal of Plant Pathology*, 109, 221–225.
- [25] Omar SA, Abd-El-Halim AZ, (1992). Fungal growth response to alfalfa (*Medicago sativa* L.) saponin extract. *Egyptian J. Appl. Sci.*, 7: 24-32.
- [26] Patel J A. (1989) Studies on wilt of Sugarcane under South Gujarath conditions. M.Sc. (Ag.) Thesis. Gujarat Agri. University,
- [27] Rahber-Bhatti, M H., (1988). Antifungal property of plant leaf decoctions for leaf rust of wheat Pak. *J. Bot.*, 20(2): 259-263.
- [28] Reddy V K and Reddy S M. (1987) . Screening of indigenous plants for their antifungal principle. *Pesticides*, 21(1987) 17-18.
- [29] Saadabi AMA, (2006). Antifungal activity of some Saudi plants used in traditional medicine. *Asian J. Plant Sci.*, 5: 907-909.
- [30] Shelef LA, (1983). Antimicrobial effects of spices. *J. Food Safety*, 6: 29- 44.
- [31] Singh J., Tripathi N.N. (1999). Inhibition of storage fungi of blackgram (*Vigna mungo* L.) by some essential oils. *Flavour Fragrance J.* 14 (1): 1–4.
- [32] Slusarenko, A. J., Patel, A. & Portz, D. (2008). Control of plant diseases by natural products: Allicin from garlic as a case study. *European journal of Plant Pathology*.
- [33] Sridhar, S.R; R.V. Rajagopal, R. Rajavel, S. Masilamani and S. Narasimhan. (2003). Antifungal activity of some essential oils. *J. Agric. Food. Chem.*, 51: 7596-7599.
- [34] Thyagaraja, N and A. Hosono. (1996). Effect of spice extract on fungal inhibition. *Lebensmittel –Wissenschaftund–Technologie.*, 29: 286-288.
- [35] Tumen, I., Fred, J.E., Carol, A.C., Jeffery, A.T. (2013). Antifungal activity of heartwood extracts from three *Juniperus* species. *BioResource*, 8 (1)12-20.
- [36] William, Q. (2008). Least toxic controls of plant diseases. *Brooklyn Botanic garden. Natural Disease Control* 11,225.
- [37] Zaker, M. and H. Mosallanejad, (2010). Antifungal activity of some plant extracts on *alternaria alternata*, the causal agent of *alternaria* leaf spot of potato. *Pak. J. Biol. Sci.*, 13: 1023-1029.